

## BACKGROUND

Phospholipase C (PLC) enzymes, comprising several families (PLC $\beta$ ,  $\gamma$ ,  $\delta$ ,  $\epsilon$ , and  $\zeta$ ), have been established as crucial signaling molecules involved in regulation of a variety of cellular functions. PLC-catalyzed formation of the second messengers, inositol 1,4,5-trisphosphate (IP<sub>3</sub>) and diacylglycerol, from phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>), constitutes one of the major cell signaling responses. IP (3) induces a transient increase in intracellular free Ca<sup>2+</sup>, while DAG directly activates protein kinase C. These second messengers provide a common link from highly specific receptors for hormones, neurotransmitters, antigens, and growth factors to downstream, intracellular targets; thus, they contribute to regulation of biological functions as diverse as cell motility, fertilization, and sensory transduction.<sup>1</sup>

Of two PLC $\gamma$  enzymes, PLC $\gamma$ 1 is ubiquitously expressed and appears to regulate a multitude of cellular functions in many tissues. It is activated in response to growth factor stimulation; in addition, its function in T-cell responses has been extensively documented. PLC $\gamma$ 2, in contrast, is most highly expressed in cells of the hematopoietic system and plays a key role in regulation of the immune response. Both PLC $\gamma$  enzymes have also been implicated in signaling events underlying aberrant cellular responses. PLC $\gamma$ 1 is critically involved in the regulation of cancer cell motility while PLC $\gamma$ 2 has been implicated in deregulation of the immune responses resembling Btk-dependent X-linked agammaglobulinemia and SLE disease in humans. It has been suggested that, in cancer cells, PLC $\gamma$ 1 could function as a key, rate-limiting, common component involved in cell motility triggered by several growth factors and integrins.<sup>2</sup>

The domain organization of PLC $\gamma$  enzymes is characterized by the insertion of a highly structured region (PLC $\gamma$ -specific array, gammaSA) between the two halves of the TIM-barrel catalytic domain common to all PLCs. The gammaSA comprises a split PH (spPH) domain flanking two tandem SH2 domains and a SH3 domain. A distinct regulatory feature of PLC $\gamma$  enzymes is that their activation is linked to an increase in phosphorylation of specific tyrosine residues (most notably within the gammaSA) by receptor and non-receptor tyrosine kinases. Upon stimulation of cells with PDGF and EGF, the SH2 domain of PLC- $\gamma$  binds to the autophosphorylated tyrosine residues of growth factor receptors, leading to tyrosine phosphorylation and activation of PLC- $\gamma$ .<sup>3</sup> In addition, activation of PLC- $\gamma$  isozymes may occur secondarily to receptor-mediated activation of phospholipase D and cytosolic phospholipase A<sub>2</sub>, which results in the production of phosphatidic acid and arachidonic acid, respectively.<sup>4</sup> It was also reported that PLC- $\gamma$  is regulated

additionally by the lipid products of PI 3-kinase. The PH domain of PLC- $\gamma$  binds to PtdIns(3,4,5)P<sub>3</sub>, and is targeted to the membrane in response to growth factor stimulation and leads to activation of PI 3-kinase causes PLC- $\gamma$  PH domain-mediated membrane targeting and PLC- $\gamma$  activation.<sup>5</sup> Furthermore, multiple protein-protein interactions (mainly mediated by SH2 domains) also contribute to activation and have an important role in localizing PLC $\gamma$  into protein complexes with different binding partners, depending on cell type and specific cellular compartments. One mode of activation that is specific for the PLC $\gamma$ 2 isozyme is direct binding to and activation by Rac. The interaction involves the spPH domain, and this activation mechanism does not require tyrosine phosphorylation.<sup>6</sup>

### References:

1. Kim, M.G. et al: Exp. Mol. Med. 32:101-9, 2000
2. Patterson, R.L. et al: Trends in Biochem. Chem. 30:688-97, 2005
3. Nishibe, S. & Carpenter, G.: Semin Cancer Biol. 1:285-92, 1990
4. Sekiya, F. et al: Chem. Phys. Lipids 98:3-11, 1998
5. Falasca, M. et al: EMBO J. 17:414-22, 1998
6. Carpenter, G. & Ji, Q.: Exp. Cell Res. 253:15-24, 1999

## TECHNICAL INFORMATION

### Source:

PLC gamma 1 Antibody is a rabbit antibody raised against a short peptide from human PLC gamma 1 sequence.

### Specificity and Sensitivity:

This antibody detects endogenous PLC gamma 1 proteins without cross-reactivity with other family members.

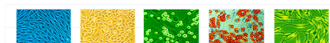
**Storage Buffer:** Rabbit IgG in phosphate buffered saline (without Mg<sup>2+</sup> and Ca<sup>2+</sup>), pH 7.4, 150mM NaCl, 0.02% sodium azide and 50% glycerol.

### Storage:

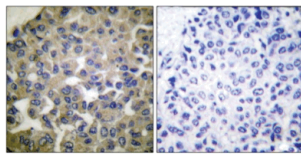
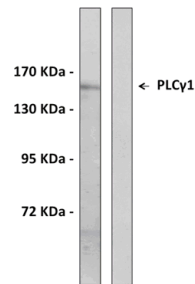
Store at -20°C for at least one year. Store at 4°C for frequent use. Avoid repeated freeze-thaw cycles.

## APPLICATIONS

Application:	*Dilution:
WB	1:500-1:1000
IP	n/d
IHC	1:50-1:100
ICC	n/d
FACS	n/d



## QUALITY CONTROL DATA



**Top:** Immunoblotting analysis of extracts from HepG2, using Anti-PLCG1 antibody. The lane on the left was treated with the Anti-PLCG1 antibody. The lane on the right (negative control) was treated with both Anti-PLCG1 antibody and the synthesized immunogen peptide.

**Bottom:** Immunohistochemistry analysis of paraffin-embedded human breast carcinoma tissue using Anti-PLCG1 antibody. Cells on the left were treated with the Anti-PLCG1 antibody. Cells on the right (negative control) were treated with both Anti-PLCG1 antibody and the synthesized immunogen peptide.

